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Connectivity clues from short-term variability in settlement and geochemical tags of mytilid mussels

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ABSTRACT

The use of geochemical tags in calcified structures of fish and invertebrates is an exciting tool for investigating 26 larval population connectivity. Evaluating these tags over relatively short intervals (weeks) may detect 27 environmental and ecological variability at a temporal scale highly relevant to larval transport and settlement. 28 We collected newly settled mussels (Mytilus californianus and M. galloprovincialis) weekly during winter/ 29 spring of 2002 along the coast of San Diego, CA, USA, at sites on the exposed coast (SIO) and in a protected 30 coastal bay (HI), to investigate temporal patterns of geochemical tags in mussel shells. Analyses of post- 31 settlement shell via LA-ICP-MS revealed statistically significant temporal variability for all elements we 32 examined (Mg, Mn, Cu, Sr, Cd, Ba, Pb and U). Despite this, our ability to distinguish multielemental signatures 33 between sites was largely conserved. Throughout our 13-week study, SIO and HI mussels could be chemically 34 distinguished from one another in 78-87% of all cases. Settlement varied between 2 and 27 settlers gram- 35 byssus⁻¹ week⁻¹ at SIO and HI, and both sites were characterized by 2-3 weeks with "high" settlement. 36 Geochemical tags recorded in early larval shell of newly settled mussels differed between "high" and "low" 37 settlement weeks at both sites (MANOVA), driven by Mg and Sr at SIO (p = 0.013) and Sr, Cd, Ba and Pb at HI 38 (p < 0.001). These data imply that shifts in larval sources or transport corridors were responsible for observed 39 settlement variation, rather than increased larval production. In particular, increased settlement at HI was 40 observed concurrent with the appearance of geochemical tags (e.g., elevated Cd), that suggest that those 41 larvae were retained in upwelled water near the mouth of the bay. Such shifts may reflect short-term changes 42 in connectivity among sites due to altered transport corridors, and influence the demography of local 43 populations. 44

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50 **1. Introduction**

The bipartite life history of many marine invertebrates and fishes 51includes a planktonic larval phase that is capable of connecting sites 52within a regional metapopulation, as well as contributing significantly 5354to spatial and temporal variability in local densities (Caley et al., 1996). Despite the achievements of researchers throughout the 19th 55and 20th centuries in investigating how larval ecology affects species 5657persistence and biodiversity maintenance (Prytherch, 1929; Thorson, 1950), there have been, until recently, severe limitations on the ability 58 to track the movement of very small, dilute larvae throughout their 59

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entire planktonic phase in the vast, dynamic ocean (Levin, 1990). 60 Levin (2006) noted that renewed vigor for tracking larvae has been 61 driven by both conservation needs [e.g., connectivity occupies a 62 central role in the design (placement of networks) and evaluation 63 (spillover and self-recruitment rates) of marine reserves (Hastings 64 and Botsford, 2006)] and methodological advances [e.g., physical- 65 biological models used to simulate larval dispersal (Cowen et al., 66 2006; Rasmussen et al., 2009)]. In particular, the discovery and 67 exploitation of environmental (geochemical) markers deposited and 68 then retained within calcified structures of larvae has allowed for the 69 reconstruction of the locations where larvae developed, and therefore 70 identification of the natal origins of settled individuals (Thorrold et al., 71 2002; Thorrold et al., 2007). The resulting insights have been 72 considerable; for example, we now understand that some populations 73 are more "self-seeding" and less demographically "open" than 74 previously expected (e.g., Almany et al., 2007). 75

The use of geochemical tags, both natural and induced, to track 76 larvae and explore connectivity remains a growth field (Campana, 77 2005; Thorrold et al., 2007). Part of the continuing challenge of these 78 studies derives from the time- and labor-intensive nature of this work, 79

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forcing datasets and conclusions to be based on few seasonal or 80 81 annual collections of settled larvae (e.g., Swearer et al., 1999; Becker et al., 2007). This is somewhat problematic given high variability of 82 83 larval dynamics over multiple spatial and temporal scales (Siegel et al., 2008). As a result, the resolution, especially temporally, of 84 geochemical tagging studies may not match up well with the scales of 85 variability that should be expected in settlement or transport 86 pathways of larvae (sensu Gaines and Denny, 1993). Indeed, the 87 88 endpoint of larval dispersal (i.e., settlement), and therefore connec-89 tivity itself, are known to vary over annual, seasonal, fortnightly and diel scales due to multiple factors such as behavior (Kingsford et al., 90 2002), upwelling relaxation (Shanks et al., 2000), the spring transition 91and wave climate (Shanks and Pfister, 2009). Thus, studies that 92explore variability in geochemical tags over a range of time scales, 93 both large and small, should add to our understanding of larval-driven 94 population connectivity (Levin, 2006; Pineda et al., 2007). It is within 95 this context that we explore and report on short-term (weekly) 96 97 variability in multielemental signatures obtained from newly arrived settlers of two common intertidal mussels. 98

In addition to changes in connectivity patterns, local environmen-99 tal fluctuations at source sites or within the water masses that larvae 100 pass through can drive variation in the geochemical tags associated 101 102 with larval shells (Strasser et al., 2008), statoliths (Zacherl, 2005) and otoliths (Gillanders, 2002) of newly settled individuals. Campana et al. 103 (2000) identified three requisites for using natural geochemical tags: 104 (1) distinct, reproducible markers among locations, (2) chemical 105characterization of all possible source groups, and (3) consistency of 106 107 signals throughout the duration of population mixing. With these rules in mind, it is also important from a logistical standpoint to 108 explore temporal variability over relatively short scales to determine 109 if the first and third of these guidelines can be satisfied in geochemical 110 tagging studies that would quantify larval connectivity. Consider, 111 seasonal (Swearer et al., 2003) and annual (Gillanders, 2002) 112 variability in multielemental signals of fish otoliths has been 113quantified as a requirement to track the nursery contribution of 114 juvenile habitats (Gillanders, 2005). Because fish occupy and then 115116 recruit from nurseries on a roughly annual basis, understanding signal variability over the scale of 0.5-1.0 years satisfies the requirements 117 presented by Campana et al. (2000). However, planktonic larval 118 durations can be much shorter than this (Thorson 1946), and **O3** 119 therefore analogous short-term studies quantifying variation in 120 121 multielemental signatures are needed, in addition to studies covering longer time scales (e.g., Zacherl, 2005). Becker et al. (2005) reported 122 temporally stable geochemical tags (Sr and Pb) in post-settlement 123 shells of mytilid mussels collected from an exposed coast site over five 124 weeks. Here, we report on an expanded mytilid dataset first used by 125126Becker et al. (2005) to further explore how temporal variability may influence, and be useful in, geochemical tagging studies. 127

Mytilus californianus and M. galloprovincialis are widely distribut-128 ed ecosystem engineers within rocky intertidal environments and 129have been valuable species for identifying the natal origins of 130131 individual larvae to estimate connectivity among sites along the 132southern California coastline (Becker et al., 2007; Rasmussen et al., 2009). These species are attractive candidates for geochemical tagging 133because: (1) each individual has a larval shell that incorporates trace 134elements and is retained after settlement, (2) they have larval 135durations between 1 and 4 weeks (Strathmann, 1987; Becker et al., 136 2007 and references therein), which are logistically manageable in 137 field experiments, and 3) they co-occur over regional and meter 138 scales. M. californianus dominate along exposed coasts and can be 139found within the outer regions of bays. Conversely, M. galloprovincialis 140are most abundant within bays but also settle along the exposed 141 coastline (Becker et al., 2005). 142

With the goal of exploring the magnitude and consequences of
short-term (weekly) variability in the geochemical tags of mytilid
mussel shells, we asked: (1) Do multielemental signatures in post-

settlement shell of mussels vary appreciably over weekly time scales? 146 If so, is this temporal variability comparable in magnitude to spatial 147 differences in geochemical signatures that might confound tracking 148 studies (and is there consistency in the elements that distinguish 149 sites)? and (2) Do geochemical tags in the portion of settled mussels' 150 shell formed during the larval phase exhibit differences based on 151 settlement date? If so, are these changes related to shifts in natal 152 sources or oceanographic conditions that affect local delivery rates of 153 settlers? 154

2. Methods

2.1. Field collections and sample preparation 156

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To investigate variability in mytilid larval settlement and geochemical tags over weekly time scales, we collected newly settled 158 mussels every week from January 25 until April 19, 2002 (13 weeks). 159 These dates overlap with typical seasonal pulses in reproduction for 160 these two species (Curiel-Ramirez and Caceres-Martinez, 2004). Our 161 collections occurred at 2 sites along the southern California coastline 162 (Fig. 1): on the most-seaward pilings of the Scripps Institution of 163 Oceanography Pier (SIO) in La Jolla, CA (N 32.87°, W 117.25°), and 164 from riprap seawalls fringing Harbor Island (HI) inside San Diego Bay, 165 CA (N 32.72°, W 117.20°). Thus, we sampled a population along the 166 exposed coast and another located in a well-flushed region (5 km from 167 the bay mouth) of a 20-km long protected bay (Chadwick and Largier, 168



Fig. 1. Location of collection sites along the southern California coastline, including one on the open coast at the Scripps Institution of Oceanography Pier (SIO) and one within a protected embayment, San Diego Bay, at Harbor Island (HI).

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1999). At both sites, collections were made at 0.3-0.7 m above the 169 170 mean lower-low tide line to minimize biases related to tidal level/ transport (Porri et al., 2007). To collect newly settled mussels, we 171 172pulled clumps of adult mussels away from underlying substrate until 3 replicate 0.5-L bags were filled. Newly settled mussels measuring 173less than 2.5 mm (\leq 2 weeks post-settlement; Coe and Fox, 1942) 174were obtained by dissecting the byssus threads that held adult masses 175together and then sorting through the byssus threads (settlement 176177 habitat for mussels) under a microscope. For each replicate 0.5-L bag, we searched for newly settled mussels for 30 min or until 30 settlers 178 were collected, whichever came first. We then dried and weighed the 179sorted byssus threads to standardize settlement rates for each site and 180week during our study (settlers gram-byssus $^{-1}$ week $^{-1}$). We also 181used a subset of these newly settled mussels to investigate spatio-182temporal variability in multielemental signals of shells, as well as 183 explore patterns of larval population connectivity. 184

We analyzed geochemical tags in shells of 181 mussels ($1.42 \pm$ 185 0.53 mm; mean \pm 1 SD), including 127 from SIO and 54 from HI. M. 186 californianus and M. galloprovincialis settlers could not be identified 187 visually. Therefore, mussel tissue samples were identified to species 188 using a molecular approach detailed in Becker et al. (2005). In short, 189 a Polymerase Chain Reaction (PCR) technique was employed using 190 191 species-specific primers targeting the 16S r-RNA subunit and 192identification of each mussel was determined from the presence and length of a PCR product. 193

Using ceramic forceps and tungsten probes to limit potential metal 194contamination, mussels were split open and flesh was removed and 195196 retained for PCR. Valves were separated and we stored the "left" valve based on the position of the dorsal apex. The "right" valve was scraped 197of debris and transferred to a clean plastic vial (if the right valve was 198 damaged, the "left" valve was used instead). Samples were leached 199overnight in 15% H_2O_2 buffered with 0.05 mol L⁻¹ NaOH, and then sonicated in 3% HNO_3^- for 5 min to further remove organics. 200 201Subsequently, shells were rinsed 3 times in Mill-Q water and then 202mounted on petrographic slides against double-stick tape using Milli-203 Q and a paintbrush. Once mussels were mounted, slides were stored 204 in a C-100 laminar flow hood until analyses. All plastic containers, 205206 glass slides, and forceps were leached in 3% HNO₃ and rinsed with Milli-O before coming in contact with mussels. 207

208 2.2. LA-ICP-MS

We analyzed the multielemental composition of mussel shells at 3 209locations: on the outer margin of dissoconch shell adjacent to the 210 dorsal apex (post-settlement shell), along the base of the prodisso-211 conch shell perpendicular to the axis of growth ("early" larval shell), 212213and on the prodissoconch shell immediately adjacent to the prodissoconch-dissoconch boundary ("late" larval shell) (Fig. 2). 214We confine this report, however, to data collected from post-215settlement shell and "early" larval shell. Because the dissoconch 216shell is deposited once mussels are settled and fixed at a site, we used 217218these data to investigate spatial variability in environmental signals 219between shell formed at SIO and HI, as well as temporal variability among weeks within both sites (question #1 above). Based on our 220221observations of laboratory-reared larvae, and growth of larvae during 7-day field outplantings (Becker et al., 2007), "early" larval shell 222223material represents the environmental conditions experienced by individual mytilids during the first week after fertilization. Therefore, 224"early" larval shell provided us an opportunity to evaluate weekly 225 variability in geochemical tags associated with the natal origin(s) or 226early larval transport corridor(s) of newly settled individuals 227 (question #2 above). 228

Shell regions were sampled using a New Wave UP 213-nm laser ablation (LA) unit. Larval and post-settlement shells were sampled by ablating a 75- μ m line with a laser output of 0.5 mJ, a scan speed of 15 μ m s⁻¹, and a burn width of 20 μ m. Experimental work by Strasser et al. (2007) demonstrated that the larval shell of softshell clams could 233 not be sampled via laser ablation without also simultaneously 234 sampling post-settlement shell. This is problematic if larval shell 235 samples are corrupted by environmental signals from the settlement 236 site (post-settlement shell) of specimens, potentially leading to 237 overestimates of self-seeding. Through visual examination of abla- 238 tions on pre-settlement mytilid larvae (Fig. 2) and careful attention to 239 Mg data collected during this study (higher concentrations in post- 240 settlement shell; Becker et al., 2007), we were confident that we could 241 fire on mussels without burning completely through larval shell (5% of 242 the larval shell data were thrown out due to concerns related to burn 243 through based on the Mg check). Furthermore, we paired post- 244 settlement and early larval shell data (X:Ca) recorded from each 245 individual mussel for regression analyses, and found that larval shell 246 data appeared largely independent of post-settlement shell (r²<0.33 247 for all eight elements we considered separately at each site). 248

Ablated shell material was transported using He gas (mixed with 249 Ar) to a Thermoquest Finnigan Element 2 double-focusing, single- 250 collector, magnetic-sector Inductively Coupled Plasma Mass Spec- 251 trometer (ICP-MS). Based on previous geochemical tagging studies in 252 this region, we sampled for the following isotopes: ²⁶Mg, ⁴⁸Ca, ⁵⁵Mn, 253 ⁶³Cu, ⁸⁸Sr, ⁶⁵Cd, ¹³⁵Ba, ²⁰⁸Pb, and ²³⁸U (Fodrie and Levin, 2008). Data ₂₅₄ processing to calculate elemental concentrations standardized to 255 calcium (X:Ca), and corrections for machine drift using NIST glass 256 (National Institute of Standards and Technology Reference Material 257 612; Pearce et al., 1996) followed Becker et al. (2005). Detection limits 258 on this instrument (3 standard deviations above background counts) 259 at the time of our analyses were: $0.02 \text{ mmol mol}^{-1}$ (Mg:Ca), 260 0.002 mmol mol⁻¹ (Mn:Ca), 0.001 mmol mol⁻¹ (Cu:Ca), 0.01 mmol - 261 mol⁻¹ (Sr:Ca), 0.004 mmol mol⁻¹ (Cd:Ca), < 0.001 mmol mol⁻¹ (Ba: 262 Ca), < 0.001 mmol mol⁻¹ (Pb:Ca) and < 0.001 µmol mol⁻¹ (U:Ca). 263 Based on ablations that produced one hundred million counts 264 of ⁴⁸Ca, the percentage of X:Ca measurements that fell below the 265 detection limits of the instrument, and the average concentration of 266 elements relative to detection limits were: Mg, 0% under detection 267 limit, average counts 68 times detection limit; Mn, 29% under 268 detection, average counts 11 times detection; Cu, 2% under detection, 269 average counts 64 times detection; Sr, 0% under detection, average 270 counts 274 times detection; Cd, 50% under detection, average counts 2 271 times detection; Ba, 0% under detection, average counts 16 times 272 detection; Pb, 1% under detection, average counts 31 times detection; 273 and U, 0% under detection, average counts 300 times detection. 274

2.3. Data analyses

2.3.1. Spatio-temporal patterns in multielemental signatures

We investigated spatial (pooling all weeks) and temporal 277 (separately for each site) differences in shell chemistry using 278 Mann–Whitney U and Kruskal–Wallis tests, respectively. Only data 279 collected from post-settlement shell were considered in these 280 analyses, and each X:Ca ratio was tested separately. Non-parametric 281 tests were employed because F_{max} tests revealed significant hetero- 282 scedasticity in shell geochemistry ($\alpha = 0.05$) for the majority of 283 elements between sites and among weeks, and log (x+1) and 284 square-root (x+1) transformations failed to reduce differences in 285 these variances.

We then used Discriminant Function Analyses (DFA) to determine 287 if SIO and HI could be characterized throughout a 13-week period by 288 distinct, multielemental signatures in post-settlement shell (Systat 9, 289 © SPSS). All DFAs were conducted in a stepwise manner, by running 290 the analysis on all element ratios, then dropping the least significant 291 variable as determined by an F-to-remove statistic. This process was 292 repeated until the F-to-remove statistic of all included element ratios 293 was>4. Based upon our visual inspections, there was an apparent 294 change in shell chemistry in the mussels collected from HI after week 295 8 (the middle of March; Fig. 3). In particular, Mn, Ba and Pb all showed 296

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Fig. 2. Images captured from the New Wave UP 213-nm laser ablation unit before (A., C.) and after (B., D.) sampling the larval and post-settlement shell of *Mytilus californianus* (A.–B.) and *M. galloprovincialis* (C.–D.). In 'before' images, larval (L) and post-settlement (Settler) shells are distinguished, and the dorsal apex is noted when visible (DA). In 'after' images, 2 laser tracks are visible and Tabeled as either "early" or "late" larval shell. Only data from the "early" larval shell and DA ablations are included in the results of this study (E., dark bars). "Early" and "late" larval shells are relative, qualitative definitions based on the primary growth axis and torsion of growing shell material observed for mytilid larvae spawned and raised in the lab (F, dark arrows).

297qualitative changes in chemical distinctness between SIO and HI following mid-March (Fig. 3). Subsequent investigations using MDS 298and SIMPER analyses (Primer 5.2.2) confirmed that shell chemistry 299was notably different at HI between the first 8 sampling weeks versus 300 the last 5 sampling weeks (unpublished data). Therefore, we 301 302 generated 3 separate DFAs to compare sites: a DFA generated with 303 data from all weeks included, a DFA with only data from the first 8 weeks, and a DFA with only data from the last 5 weeks. Cross-304validation of each DFA model was achieved by re-classifying each 305 sample using a jackknife method, and comparing observed classifi-306 cation successes to the average of six replicate trials in which the 307 collection site of individual mussel settlers was randomly assigned 308 (White and Ruttenberg, 2007). 309

310 2.3.2. Temporal patterns in geochemical tags of early larval shell

We also employed DFA to evaluate the coherence (spatiotemporal) of geochemical tags in early larval shell among settled mussels at SIO or HI during our 13-week study. As before, this DFA was run in a stepwise manner, dropping element ratios until all F-toremove values were greater than 4, cross-validating the model using the jackknife method, and comparing our observed classification 316 success against the average of six replicate trials with mussel 317 collection sites randomly reassigned. Because we were only interested 318 in gauging the within- and between-site similarities of geochemical 319 tags within early larval shell, rather than attempting to explicitly 320 define the natal origins of settled larvae, we did not employ additional 321 statistical approaches on these data such as Markov Chain Monte 322 Carlo methods (White et al., 2008). 323

Settlement rates of mytilid mussels at SIO and HI were defined by a 324 few weeks with strong pulses of newly arrived larvae interspersed 325 among weeks with "low" background settlement levels. We differen-326 tiated "high" settlement phases as weeks with settlement greater than 327 three standard deviations above mean settlement at that site (after 328 removing the week in question from the calculation of mean 329 settlement). As a result, the first, third and ninth weeks at SIO were 330 deemed "high" settlement phases, while at HI the tenth and twelfth 331 weeks were considered "high" settlement phases (Fig. 4). 332

To test if there were distinct natal or transport signatures in early 333 larval shell for settlers between settlement phases, we used separate 334 MANOVA (StatView 5.0.1, © SAS) analyses for each site to compare 335

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Fig. 3. Temporal patterns of multielemental signatures in post-settlement mussel shell collected from Scripps Pier (SIO) and Harbor Island (HI). Mg:Ca (A), Mn:Ca (B), Cu:Ca (C), Sr:Ca (D), Cd:Ca (E), Ba:Ca (F), Pb:Ca (G) and U:Ca (H). Gray vertical bars indicate an apparent shift in environmental conditions following week 8. Cd, Ba, Pb and U were included in a DFA to compare multielemental signatures in post-settlement shell between sites for the entire sampling period; Ba, Pb and U were included in a DFA for only the first eight weeks; and Mg, Sr, Cd and U were included in a DFA for only the last five weeks.



Fig. 4. Settlement of mytilid mussels (settlers gram-byssus-thread⁻¹) during the winter and spring of 2002 at Scripps Pier (SIO) and Harbor Island (HI). Weeks classified by "high" settlement events (>3 SD above mean settlement) are denoted by *H* (SIO) or H (HI).

early larval shell chemistry of individual mussels collected between 336 "low" or "high" settlement phases (with all weeks pooled between 337 phases). For each site, only elements that remained in an exploratory 338 DFA to compare "high" and "low" settlement phases were included in 339 the MANOVA. If early larval shell chemistry was not different between 340 "high" and "low" settlement phases at a site, this would suggest that 341 changes in settlement rates were the result of increased larval 342 production or survivorship. If early larval shell chemistry was 343 different between "high" and "low" settlement phases, this would 344 indicate that changes in larval sources, or the water masses through 345 which larvae passed during early development (perhaps interacting 346 with larval production or survivorship), played some role in 347 regulating the observed settlement rates at SIO or HI. 348

An alternative hypothesis for why we might observe significant 349 differences in early larval shell chemistry between "high" and "low" 350

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settlement phases would be that there are changes in environmental 351 352 conditions among all weeks, rather than anything specifically related to the observed settlement patterns. To evaluate this hypothesis, we 353 354 randomly selected three (SIO) or two (HI) weeks and compared the geochemical tags in early larval shell of settlers collected during those 355 randomly selected weeks to settlers from all other weeks. This was 356 repeated six times, and we then compared the results of each 357 MANOVA result ("high/low" and the six "random/all other week" 358 359 tests) to provide a more complete context for our statistical inferences. As an additional output of MANOVA testing, element-by-360 element ANOVAs comparing settlement phases were run. Data 361 transformations were not required to reduce differences in variances 362 between groups. Because each statistical test we conducted applied to 363 separate and easily distinguishable hypotheses, we made no correc-364 tions to experiment-wise alpha for either the parametric or non-365 parametric tests we conducted (Moran, 2003). 366

367 3. Results

368 3.1. Weekly settlement

Settlement ranged between 2 and 22 settlers gram-byssus⁻¹ 369 370 week⁻¹ at SIO and 2 and 27 settlers gram-byssus⁻¹ week⁻¹ at HI (Fig. 4). As noted above, the first, third and ninth collection weeks at 371 SIO were qualified as relatively "high" settlement phases, while the 372 tenth and twelfth weeks were considered "high" settlement phases 373 at HI. Genetic identification of the specimens analyzed via LA-ICP-374 375 MS revealed that all of the settlers at HI and 9% (n = 11) of settlers at SIO were *M.* galloprovincialis. The remaining 91% (n=109) of 376 settlers at SIO were M. californianus. 377

378 3.2. Spatio-temporal patterns in multielemental signatures

There were significant differences ($\alpha = 0.05$) in the elemental 379 signatures of post-settlement shell (X:Ca) between SIO and HI for Mg, 380 Mn, Sr, Cd, Ba, Pb and U (Table 1, Fig. 3). These differences were most 381 apparent during the first 8 weeks of the study (Jan 26-March 15). 382 383 During this interval Mg was, on average, elevated in mussel shell at HI over SIO by a factor of 2; Mn concentrations ranged between 2 and 20 384 fold greater at HI than at SIO; average Cu concentrations were nearly 385 3-times higher in mussel shells collected at HI; Ba was 2-6 fold higher 386 387 at HI than at SIO; Cd was up to10-times more elevated in HI shells (when measures were above detection limits); and Pb was more 388 abundant in shells from HI (Fig. 3). Conversely, Sr concentrations 389

were typically higher in the post-settlement shells of mussels $_{390}$ collected at SIO. During the last 5 weeks we collected mussels, these $_{391}$ X:Ca differences between sites tended to decrease, or even exhibit a $_{392}$ phase change in the case of Cu and Sr (Fig. 3). At SIO, significant $_{393}$ (p<0.05) temporal variability was observed for all elements except Pb $_{394}$ (Table 1). At HI, Mn, Cd, Ba, Pb and U concentrations varied $_{395}$ significantly (p < 0.05) in post-settlement shell among weeks $_{396}$ (Table 1).

During the winter and spring of 2002, the multielemental 398 signatures of post-settlement mussel shells collected at SIO and HI 399 could be distinguished from one another using DFA with 80% accuracy 400 (compared to only 55% during random assignment trials). Notably, all 401 11 of the M. galloprovincialis settlers at SIO were correctly identified to 402 their collection site, indicating that the discrimination between SIO 403 and HI was a true site distinction rather than just a species comparison 404 (i.e., that spatial gradients in geochemical tags contributed more 405 toward our results than did potential [expected] species differences). 406 DFA accuracy was 87% for mussels collected during the first eight 407 sampling weeks (compared to 52% random) and 78% for mussels 408 collected during the last five weeks (compared to 53% random) 409 (Table 2). Regardless of the sampling interval, classification success 410 was higher at SIO than at HI by 7-30%. Although DFA accuracy was 411 conserved across the three sampling intervals, the elements that 412 drove DFA algorithms varied notably. For the entire 13-week study, 413 Ba, U, Pb and Cd (in decreasing relative importance) drove differences 414 between SIO and HI. Ba, Pb and U were used to discriminate sites 415 during the first 8 weeks, while Cd, Sr, Mg and U were used in the DFA 416 during the last 5 weeks (in decreasing relative importance). 417

3.3. Temporal patterns in geochemical tags of early larval shell 418

We were able to extract early larval shell geochemical data from 419 151 individual mussels, and observed distinct early larval tags 420 between the settlers at SIO and HI based on DFA. The mean $(\pm 1 421$ standard error) score of the lone DFA algorithm used to distinguish 422 individuals between sites was -1.099 ± 0.156 (SE) for settlers 423 collected at HI, while the mean score for individuals collected at SIO 424 was 0.431 ± 0.096 (SE). Settlers at HI were defined by early larval 425 shell typically more enriched with Ba, while settlers at SIO generally 426 exhibited higher concentrations of Mg and U. Overall, geochemical 427 tags in early larval shells collected between the two sites could be 428 distinguished using a jackknife approach in 83% of cases as "SIO type" 429 or "HI type", compared to only 57% in trials with collection site 430 randomized among specimens (Table 2).

t1.1 Table 1

Summary table of X:Ca ratios in mytilid mussel post-settlement and larval shells collected from the Scripps Institution of Oceanography Pier (SIO) and Harbor Island riprap seawall within San Diego Bay (HI).

t1.2 t1.3		Mg:Ca	Mn:Ca	Cu:Ca	Sr:Ca	CdCa	Ba:Ca	Pb:Cata	U:Ca
t1.4	Post-settlement shell concentration (mmol mol ⁻¹)								
t1.5	SIO (n120)	1423 ± 0.058	0.056 ± 0.009	0.069 ± 0.005	2.238 ± 0.071	0.006 ± 0.001	0.010 ± 0.001	0.016 ± 0.002	$0.003\pm0.00~\text{L}$
t1.6	HI $(n = 5 L)$	1.670 ± 0.090	0421 ± 0132	0.079 ± 0.009	1.890 ± 0.100	0.012 ± 0.005	0.023 ± 0.001	0.096 ± 0.026	0.001 ± 0.0001
t1.7	Site comparison (Mann–Whitney U)								
t1.8	U	2486	992	3142	2652	1722	1876	1390	2777
t1.9	z-value	-2.924	-7556	-0.890	-2.409	-5.293	-4.815	-6.322	-2.022
t1.10	p-value	0.004	< 0.001	0.374	0.016	<0.001	-0.001	<0.001	0.043
t1.11	Temporal comparison (Kruskal–Wallis)								
t1.12	SIO	12	12	12	12	12	12	12	12
t1.13	df	36.517	25 <mark>,</mark> 384	43.486	23.449	31.310	31,283	15.980	59.535
t1.14	p-value	< 0.001	0.013	< 0.001	0.024	0.002	0.192	< 0.001	
t1.15	HI								
t1.16	df	12	12	12	12	12	12	12	12
t1.17	Н	17.780	26.506	19.168	11.709	25.555	22,231	29.436	2.5.833
t1.18	p-value	0.123	0.009	0.085	0.469	0.012	0.5	0.003	0.011
t1.19	¹⁹ Larval shell concentration (mmol mol ⁻¹)								
t1.20	SIO (n 108)	$0267\pm0.02.5$	0.102 0.022	0.016 ± 0.002	3.319 ± 0.080	0.002 ± 0.001	0.010 ± 0.001	0.037 ± 0.005	0.003 ± 0.001
t1.21	HI (n=43)	0292 ± 0.051	0.400 ± 0.133	0.036 ± 0.010	2.745 ± 0.146	0.122 ± 0.116	0.016 ± 0.002	0.094 ± 0.024	0.001 ± 0.001

t1.22 Included are the effects of site (weeks pooled) and time (among weeks within a site) on post-settlement shell chemistry based on non-parametric testing.

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t2.1 Table 2

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Classification success (jackknifed) of DFA algorithms used to distinguish: 1) multielemental signals in post-settlement shell between mussels collected at Harbor Island (HI) within San Diego Bay and at Scripps Pier (SIO) along the open coast, or 2) geochemical tags in early larval shell of settled mytilid mussels specimens collected at HI and SIO (used to infer larval dispersal).

	Predicted	Predicted		Classification success %		
	HI	SIO	Correct	Random		
Post-settlement	shell: all weeks					
Actual						
HI	30	21	59	43		
SIO	14	106	89	65		
Total	444	127	80	55		
Post-settlement	shell: last 8 weeks					
Actual						
HI	20	8	71	49		
SIO	7	76	93	53		
Total	27	84	87	52		
Early larval she	ell: last 5 weeks					
HI	17	6	74	52		
SIO	7	30	81	54		
Total	24	36	78	53		
Early larval she	ell: natal origins					
	Natal orig	Natal origin		Larval trajectory %		
	HI	SIO				
	"type"	"type"	Local <mark>"</mark> type"	Random		
Settlement Site	2					
HI	39	4	91	48		
SIO	21	87	81	60		
Total	60	91	83	57		

Rows list the collection site of specimens, while columns register the predicted collection site (for post-settlement shell) or natal signature (for early larval shell) of individuals based on shell chemistry entered in to a DFA model. For post-settlement shell, classification successes are presented for the entire sampling period, during only the for the during of previous devices and the shell she to be the formula of previous devices.

t2.31 the first 8 weeks of sampling and during only the last 5 weeks of sampling.

Within each site, the geochemical tags in early larval shells were 432 also distinct between "high" and "low" settlement phases (Table 3). 433 434 Mg and Sr were included in MANOVA analyses for SIO and revealed significant differences between settlement phases (p = 0.013). Both 435436 Mg (p = 0.018) and Sr (p = 0.119) were enriched in the early larval shells of settlers during "high" settlement phases (Fig. 5). Conversely, 437 comparisons between geochemical tags of early larval shell from 438 439 settlers at SIO during 3 randomly selected weeks and all others 440 revealed non-significant results (n=6 random trials, average)441 p = 0.393, all p > 0.2). At HI, Sr, Cd, Ba, and Pb were included in the MANOVA and indicated a significant difference in the geochemical 442

t3.1 **Table 3** Effect of settlement phase ("low"

t3 11

Effect of settlement phase ("low" versus "high") on the geochemical tags within early larval shell of settled mytilid mussels at the Scripps Pier (SIO; 2 elements) and at Harbor Island within San Diego Bay (HI; 4 elements) based on MANOVA.

t3.2 t3.3	Elements	SIO	HI	
t3.4		Mg, Sr	Sr, Cd, Ba, Pb	
t3.5	MANOVA score	0.087	0.434	
t3.6	df	2	2	
t3.7	df-residual	108	43	
t3.8	F-value	4.563	9.335	
t3.9	p-value	0.013	< 0.001	
t3.10	p-value (random)	0.393	0.494	

Also included are the average MANOVA results for 6 trials in which settlers during three (SIO) or two (HI) randomly selected weeks were compared to settlers from all other weeks.



Fig. 5. Elemental concentrations (X:Ca) in early larval shell of mytilid mussels collected during "low" and "high" (>3 SD above mean settlement) settlement phases at Scripps Pier (SIO) and Harbor Island (HI). For each element used in MANOVA_testing, element-by-element comparisons between recruitment phases were generated via t-tests, with significant results denoted by *(p<0.05) and **(p<0.001).

tags of early larval shell between "high" and "low" settlement phases 443 (p<0.001). Sr (p=0.049) and Cd (p<0.001) were enriched in early 444 larval shells of "high" phase settlers, while both Ba (p=0.011) and Pb 445 (p=0.029) concentrations were lower in those individuals (Fig. 5). 446 **Comparisons of** geochemical tags between settlers collected in 2 447 randomly selected weeks versus all other weeks at HI were not 448 significant (n=6 random trials, average p=0.494, all p>0.2). 449

4. Discussion

We investigated how temporal variability in the geochemical tags 451 of mussel shells may influence and aid estimates of larval connec- 452 tivity. For logistical reasons, we tested if temporal variability in shell 453 chemistry at our two collection sites could obscure site-specific 454 signatures over a time scale relevant for exploring larval connectivity 455 (~ weekly). Early in our study, differences in post-settlement shell 456 geochemistry reflected known environmental gradients between our 457 one bay and one open coast site. Previous studies in this region on 458 crab (DiBacco and Levin, 2000), mussels (Becker et al., 2005; Becker 459 et al., 2007) and fish (Fodrie and Herzka, 2008; Fodrie and Levin, 460 2008) have all reported elevated concentrations of Mn (redox cycles 461 in muddy sediments), Cu (boat paints), Ba (salinity fractionation) 462 and Pb (pollution) in the hard parts of organisms developing within 463 San Diego Bay relative to the exposed coast. Previously, Becker et al. 464 (2005) reported limited variability among weeks in multielemental 465 signatures of post-settlement shell at SIO during January 09-466 February 12, 2002. Our expanded analyses of mytilid mussels from 467 January 25 to April 19, 2002, confirmed the findings of Becker et al. 468 (2005), but also suggested that all elements we examined in post- 469 settlement shell (particularly Mn, Cu, Cd, Ba, and Pb) could be 470 relatively distinct between our two collection sites over several 471 weeks (January 25-March 15), and then at one of the sites we studied 472 (HI), quickly shift to lower concentrations for several more weeks 473 (March 15-April 20). Generally, this change after mid-March 474 resulted in multielemental signatures at HI during the last five 475 weeks that were quantitatively more similar to those observed at SIO 476 throughout the 13-week study. 477

Often, changes in bay-ocean exchange due to either wind or tidal 478 forcing, or changes in the amount of fresh water runoff (i.e., rainfall) 479 are invoked to explain temporal variability in shell/otolith geochem-480 istry within estuarine systems (Gillanders and Kingsford, 1996). 481 However, *post hoc* examination of wind data from the Coastal Data 482 Information Program station 73 at SIO (http://cdip.ucsd.edu), tide 483 data from the National Oceanic and Atmospheric Administration buoy 484 station 9410230 in La Jolla, CA (http://tidesandcurrents.noaa.gov), 485 and rainfall data at Lindbergh field in San Diego, CA (http://cdec. 486 water.ca.gov), during January-April of 2002 reveal no clear explanation for the shift in multielemental signatures in post-settlement shell 488

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at HI away from a "bay-type" signature following March 15. Throughout our study, winds were typically mild ($<4 \text{ m s}_1^{-1}$), spring and neap tides were experienced during each month, and precipitation (i.e., runoff) was actually higher during March_April (19.6 mm) than during January_February (12.5 mm).

Despite the temporal variability we recorded, our ability to 494 generate distinct chemical tags from post-settlement shell between 495SIO, on the open coast, and HI, within a protected bay, was largely 496 497 unhampered. Even during the last five weeks of our study, when signals at SIO and HI appeared to converge, multivariate analyses 498 499(DFA) were able to tease apart unique multielemental signatures in post-settlement shell and allow for the correct identification of 500collection site for individual mussels 78% of the time (compared to 50150280% and 87% for the entire 13 weeks and first 8 weeks, respectively). Thus, despite some variability among weeks, our data suggest that it is 503possible at the scale of a single bay site versus a single exposed coast 504site to satisfy at least two of the requirements Campana et al. (2000) 505 listed for employing geochemical tags to track larvae: (1) distinct, 506reproducible markers among locations, and (2) chemical character-507ization of all sources. 508

Importantly, our data indicate that species effects did not play a 509major role in our findings even though 100% of the settlers at HI were 510511 M. galloprovincialis, while 90% of the settlers at SIO were M. californianus. All 11 M. galloprovincialis we analyzed via LA-ICP-MS 512that settled at SIO were correctly identified to their collection sites 513based on post-settlement shell geochemistry. Thus, multielemental 514discrimination between SIO and HI was a true site distinction rather 515516than just a species comparison. This is not to say that species difference does not exist for certain X:Ca shell concentrations or 517multielemental geochemical tags, but that in this study system those 518differences were relatively minor when compared to spatial gradients 519520in shell chemistry.

521Our data also identify points of caution regarding temporal variability in multielemental signatures. We found that different 522elements defined the geochemical tags at SIO and HI shell during the 523first 8 weeks (Ba, PB and U) and the final 5 weeks (Cd, Sr, Mg and U). 524525 This indicates, to a manageable degree, that the third requirement advised by Campana et al. (2000) is more difficult to meet: temporal 526 consistency of chemical signals. Clearly, it is important to quantify 527site-specific, reference signatures indicative of natal origins at the 528time larval structures are forming and over a time scale appropriate 529530for a typical planktonic larval stage (i.e., days-weeks). For instance, using a geochemical atlas generated in late March to determine the 531 natal origin of larvae developing during early March (or vice versa) 532during 2002 could have generated misleading results (albeit based on 533post-settlement shell data). We also recognize that we only collected 534535settlers at two sites, and this limits our ability to negate temporal variability as a concern for geochemical tagging studies. For instance, 536two sites within San Diego Bay might become completely indistin-537guishable, or even mistaken for one another, given the magnitude of 538geochemical variation we observed at HI. Ultimately, however, we 539540expect there are identifiable "regions" (25-100 km) over which 541relatively stable, characteristic elemental signals can be used to explore larval connectivity (e.g., Becker et al., 2005; Zacherl, 2005; 542Carson et al., 2008). 543

Throughout our 13-week study, the early larval chemical signa-544545tures of newly settled mussels collected at either SIO (mainly M. californianus) or HI (M. galloprovincialis) were distinguishable from 546each other as "SIO type" or "HI type" (83% overall classification 547success). Without identifying the natal origin(s) of these larvae, we 548could hypothesize that most (91%) of settlers at HI had a distinct natal 549source from that of most (81%) settlers at SIO (Table 2). Specifically, 550we found that early larval shells of setters at HI were relatively 551enriched with Ba (indicative of bay environments; DiBacco and Levin, 5522000; Becker et al., 2005) while Mg was more enriched in the early 553554 larval shells of SIO settlers (indicative of exposed environments; DiBacco and Levin, 2000; Fodrie and Herzka, 2008). These results 555 suggest high self-seeding rates at a coarse habitat level for the HI and 556 SIO populations. This is predictable given the distribution of *M*. 557 californianus and *M*. galloprovincialis, although DiBacco and Levin 558 (2000) did find considerable exchange of crab zoea between San 559 Diego Bay and the exposed coast, while Becker et al. (2007) reported 560 divergent scenarios for *M*. californianus (little exchange) and *M*. 561 galloprovincialis (moderate exchange). 562

Without a detailed chemical atlas of potential source populations 563 (i.e., we only sampled two sites), we hesitate to go further and 564 quantitatively estimate exchange rates between and among bay and 565 exposed coast populations. We also have reasons to qualify our 566 classification of higher Ba and lower Mg in early larval shell as a 567 signature indicative of bay environments, as these expectations are 568 largely drawn from data we extracted from post-settlement shell 569 (although confirmed in other studies). Becker et al. (2007) discussed 570 the differences in mineralogy between post-settlement (aragonite/ 571 calcite mix) and larval (mostly aragonite) shells of mussels that 572 affected Sr and Mg uptake rates, and subsequently relied on larval 573 outplanting as the best approach for generating a chemical atlas of 574 potential source populations for larval tracking.

The geochemical tags in early larval shell of mussels during "high" 576 and "low" settlement phases were distinct at both SIO and HI. These 577 data may suggest that changes in reproductive output or larval 578 survival alone did not drive the observed variability in settlement 579 rates. Rather, we hypothesize that newly settled mussels carried a 580 chemical marker that suggested changes in (1) larval sources or (2) 581 the water masses in which developing larvae passed through (as we 582 sampled approximately 1 week of shell growth during our ablations; 583 Fig. 2), also contributed to settlement variability. 584

The data from HI were particularly intriguing. At HI, source 585 signatures in larval shells of M. galloprovincialis appeared more 586 influenced by exposed coast conditions during "high" settlement 587 phases than during "low" settlement phases (i.e., higher Sr, lower Ba 588 and lower Pb). Perhaps most tellingly at HI, Cd concentrations in larval 589 shell were ~100 times more enriched during "high" settlement weeks 590 than during "low" settlement weeks. Cadmium has previously been 591 shown to be a clear indicator of upwelling in the waters adjacent to 592 San Diego Bay (seawater concentrations elevated by 50-fold relative 593 to non-upwelling conditions; Segovia-Zavala et al., 1998), and is 594 dependably recorded in *M. californianus* as an indicator of upwelling 595 along the West Coast (Lares and Orians, 1997). Recently, Levin (2006) 596 noted that "evaluat[ing]... larval movements through upwelling 597 zones, oxygen minima, turbidity plumes, warm or cold eddies, or 598 salinity fronts" is among five important directions in which 599 geochemical tags should be applied. With this in mind, we consider 600 briefly how our larval shell data might lead to future, more rigorous 601 studies that evaluate the role upwelling plays in determining 602 transport corridors and realized larval population connectivity for 603 mytilid mussels in this region. 604

In particular, we hypothesize that changes in local oceanographic 605 conditions near San Diego Bay (i.e., upwelling) affected settlement 606 rates of *M. galloprovincialis* at HI based on our analyses of early larval 607 shell. Upwelling (Pineda, 1991) and retention zones in the lees of 608 headlands (Mace and Morgan, 2006) have strong effects on dispersal 609 and settlement of larvae for many nearshore species. Roughan et al. 610 (2005) reported isolated upwelling during early April, 2003, in the lee 611 of Point Loma, immediately adjacent to the mouth of San Diego Bay 612 (Fig. 1), following the offshore divergence of the dominant southerly 613 flow as it passed this headland. It is plausible, although ultimately 614 untested, that similar oceanographic conditions occurred intermit- 615 tently during our study, and that some larvae were entrained in 616 upwelled water in the lee of Point Loma. This is supported by the 617 change in post-settlement shell chemistry at HI following week 8, 618 assuming that some of the upwelled coastal water entered San Diego 619 Bay. If this water mass retained M. galloprovincialis larvae near San 620

Diego Bay and decreased offshore wastage, or increased survivorship 621 622 because of (a) enhanced feeding opportunities for larvae, or (b) 623 reduced predation pressure relative to within the Bay (DiBacco and **O4** 624 Levin 2008), this could explain the settlement peaks we recorded that were associated with a geochemical tag indicative of upwelling 625 (elevated Cd). Although upwelled water would eventually advect 626 offshore (Roughan et al., 2005), upwelling is not necessarily a barrier 627 to nearshore retention for larval bivalves (Shanks and Brink, 2005; 628 629 Shanks and Shearman, 2009), particularly in this system where upwelling occurs over just a few kilometers (Roughan et al., 2005). 630

631 Variability in pre-recruitment dynamics (dispersal pathways) is 632 known to drive large fluctuations in population size and age structure 633 for many marine species. For instance, Gaines and Bertness (1992) 634 found that shifting transport corridors (retention versus export) near Narragansett Bay, Rhode Island, was the mechanism behind variable 635 recruitment. Specifically, high settlement occurred when the flushing 636 time (forced by riverine input) of the bay was more than 25 days and 637 larval retention was high, and this only occurred in 3 of 9 years during 638 their study. Similarly, Kraus and Secor (2005) demonstrated that 639 during most years, recruitment pulses of white perch in Chesapeake 640 Bay were mainly from freshwater nurseries. However, in years that 641 produced the dominant year-classes of the population, recruitment 642 643 pulses came mostly from brackish nurseries. Locally, Rasmussen et al. 644 (2009) showed that relatively small changes in the wind field along the San Diego coast (and more specifically, uncertainty in the 645 dynamics of wind-driven circulation near a geomorphologically 646 complex shoreline) could significantly affect measures of regional-647 648 scale connectivity for a passive tracer. Using a bio-physical model of "fish" larval dispersal along an idealized coastline, Siegel et al. (2008) 649 demonstrated that episodic events driven by interactions between 650 larval life histories and complex coastal circulation would result in 651 652 unpredictable settlement even in the most homogeneous environments. Therefore, it follows that larval connectivity would be 653 654inherently stochastic and highly temporally variable. Taken together, 655 these data on fish and invertebrates, in combination with our data, highlight the importance of incorporating measures of variability in 656 estimates of population connectivity, as larval ecology cannot be well 657 658 described by mean conditions (Siegel et al., 2008). Thus, we conclude that investigating variability in the geochemical tags of larval hard 659 parts over a range of scales [from diel (i.e., internal bore warm fronts) 660 to decadal (oceanographic/reproductive cycles related to El Niño 661 Southern Oscillation)] remains an exciting avenue in the development 662 of methods for exploring larval ecology and population connectivity 663 (Pineda et al., 2007; Thorrold et al., 2007). 664

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